Enhanced Disease Resistance in Plants



TECHNOLOGY HIGHLIGHTS

- BTI and Cornell scientists have identified the *Ptr1* gene in *Solanum lycopersicoides*, a wild relative of tomato
- Domesticated tomato varieties lack a functional copy of the Ptr1 gene
- Tomato lines expressing the S. lycopersicoides Ptr1 gene remained free of bacterial speck disease in the field during a natural outbreak
- The S. lycopersicoides Ptr1 gene confers resistance to major bacterial pathogens causing wilt, speck and spot disease in plants
- Traditional plant breeding or genetic engineering approaches can be used to transfer or restore Ptr1 functionality in tomatoes and possibly other crops such as eggplant

EFFECTIVENESS OF THE PTR1 TECHNOLOGY



Ptr1 confers resistance to bacterial speck disease.

Plants were inoculated with 1 x 10^4 cfu/mL NYS-T1 race 1 *P. syringae* pv. tomato strain, which expresses the AvrRpt2 protein.

Photographs were taken 7 days after inoculation.

LICENSING OPPORTUNITIES



Genetic engineering Exclusive licenses are

Exclusive licenses are available on a species per species basis

Masker-assisted plant breeding

Non-exclusive licenses available

COLLABORATION/R&D OPPORTUNITIES



BTI and the Martin lab will consider proposals for company-sponsored research or participation in SBIR or STTR grants. An area of particular interest is the application of the *Ptr1* technology to species other than tomatoes.

INTELLECTUAL PROPERTY



ENHANCED DISEASE RESISTANCE IN PLANTS

U.S. Application 16/916,757 Status: pending Inventors: Greg Martin, Ari Feder, Sarah Hind, Samantha Mainiero, and Diana Carolina Mazo-Molina Assignees: Boyce Thompson Institute and Cornell University

Key Facts About The Technology

BACKGROUND INFORMATION

Bacterial speck causes severe necrotic lesions on different parts of the plant, affecting fruit yield and quality, requiring chemical treatment to avoid significant economic losses.

The Pto/Prf genes confer genetic resistance to race 0 strains of *P. syringae* by coding for a serine/threonine cytoplasmic kinase and a nucleotide-binding leucinerich repeat (NLR) protein. These proteins form a complex that recognizes the bacterial type III effectors AvrPto or AvrPtoB

However, the widespread use of the Pto/Prf genes in the 1980s has led to the emergence of race 1 strains of *P. syringae* lacking AvrPto and AvrPtoB

Ptr1, by detecting the presence of the bacterial protease AvrRpt2 present in race 1 strains, confers strong resistance against bacterial speck. $^{[1,2]}$ Ptr1 also recognizes the *P. syringa* effectors HopZ5, AvrRpm1 and AvrB. $^{[3]}$

Bacterial spot disease is caused by members of the Xanthomonas genus, By recognizing the effector AvrBst, Ptr1 also confers resistance to bacterial spot disease.^[3]

BROAD RESISTANCE TO BACTERIAL PATHOGENS





Ptr1 confers resistance to bacterial wilt disease.

Plants were soil drench-inoculated with 50 mL of 10⁸ cfu/mL of *R. pseudosolanaceraum* CMR15. which expresses AvrRpt2 homolog RipBN. Photographs were taken 13 days after inoculation.

Bacterial wilt is caused by members of the Ralstonia genus. Ptr1 was first found to recognie RipBN present in strains of phylotype III.^[3] Ptr1 also recognizes the effector RipE1, commonly found in all Ralstonia phylotypes ^[4].

Ptr1 has been proposed as a key component of a stacking strategy for durable resitance to a broad range of bacterial diseases.^[5]

HOW TO USE PTR1 IN TOMATO

Genetic engineering:

A Ptr1 gene insertion or the correction of the pseudogene present in cultivated tomato are possible.

Marker-assisted plant breeding:

Introgression of the *Ptr1* gene into tomato varieties is also possible.

HOW TO USE PTR1 IN OTHER PLANTS

Use in Solanacea:

Genomic data indicates that Ptr1 may have been lost across tomato and eggplant cultivars. Analyzed accessions of tobacco, pepper and potato presented a functional copy of Ptr1 [2]. Some cultivated varieties/lines however may have lost Ptr1 during domestication.

Use in other crops:

Research is needed to determine whether the introduction of *Ptr1* could confer resistance in plants outside of the *Solanaceae* family.

References

[1] The Ptr1 Locus of Solanum lycopersicoides Confers Resistance to Race 1 Strains of Pseudomonas syringae pv. tomato and to Ralstonia pseudosolanacearum by Recognizing the Type III Effectors AvrRpt2 and RipBN. Mazo-Molina et al., Mol Plant Microbe Interact. 2019 32(8):949-960. [2] Ptr1 evolved convergently with RPS2 and Mr5 to mediate recognition of AvrRpt2 in diverse solanaceous species. Mazo-Molina et al., Plant J. 2020 103(4):1433-1445.

[3] Ptr1 and ZAR1 Immune Receptors Confer Overlapping and Distinct Bacterial Pathogen Effector Specificities. bioRxiv preprint doi: https://www.biorxiv.org/content/10.1101/2022.05.16.492216v1.

[4] The Core Effector RipE1 of Ralstonia solanacearum interacts with and cleaves Exo70B1 and is recognized by the Ptr1 immune receptor. Tsakari et al., bioRxiv preprint doi: https://www.biorxiv.org/content/10.1101/2022.08.31.506019v3.

[5] Tackling Multiple Bacterial Diseases of Solanaceae with a Handful of Immune Receptors. Kim et al., Hortic Environ Biotechnol. 2022 63:149-160.

MEET OUR FACULTY/INVENTOR

Greg Martin is Professor in the School of Integrative Plant Science, Plant Pathology and Plant-Microbe Biology Section at Cornell University. Greg Martin is also a member of the National Academy of Sciences.

The Martin lab at BTI studies the molecular basis of bacterial infection processes, plant disease susceptibility, and plant immunity using biochemistry, bioinformatics, cell biology, forward and reverse genetics, genomics, molecular biology, plant breeding, plant pathology and structural biology.





BTI's mission: To advance and communicate scientific discovery in plant biology to improve agriculture, protect the environment, and enhance human health.

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